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Application of Proteomic Tools in Modern Nanotechnological Approaches Towards Effective Management of Neurodegenerative Disorders

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Abstract: Neurodegeneration is the progressive loss of structure or function of neurons leading to neuronal death, usually associated with ageing. Some of the common neurodegenerative disorders include Alzheimer's disease, Parkinson's disease, Creutzfeldt-Jakob disease, and Huntington's disease. Due to recent advancements in high-throughput technologies in various disciplines such as genomics, epigenomics, metabolomics and proteomics, there has been a great demand for detection of specific macromolecules such as hormones, drug residues, miRNA, DNA, antibodies, peptides, proteins, pathogens and xenobiotics at nano-level concentrations for in-depth understanding of



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disease mechanisms as well as for the development of new therapeutic strategies. The present review focuses on the management of agerelated neurodegenerative disorders using proteomics and nanotechnological approaches. In addition, this review also highlights the metabolism and disposition of nano-drugs and nano-enabled drug delivery in neurodegenerative disorders.

Keywords: Disposition, nano-applications, nano-drugs, nano-techniques, nanotechnology, neurological disorders, proteomics.

1. INTRODUCTION

The term neurodegeneration is generated by a combination of two different words, 'neuro' meaning 'nerve cells' and 'degeneration' meaning 'progressive damage'; and can be applied to various conditions that result in continual loss of neuronal structure and function finally leading to neuronal death [1]. Common neurodegenerative disorders (NDDs) include Alzheimer's disease (AD), Parkinson's disease (PD), Creutzfeldt-Jakob disease, frontotemporal dementia and Huntington's disease [2]. Progressive accumulation and aggregation of proteins like tau, α -synuclein and amyloid- β (A β) have been reported to be involved in the gradual development of various NDDs [3, 4]. Unfortunately, to our knowledge, there is no single drug available that can halt or even slow down the progress of brain degeneration caused by NDDs. Apart from adverse effects on human health, NDDs have been reported to exhibit significant associations with other chronic diseases like cancer, diabetes and cardiovascular diseases [4-8]. Many different approaches have been proposed and utilized to develop cure of these chronic diseases [9-13], but full proof treatment options are still obscure. This calls for an urgent need to develop accurate and informative diagnostic tests as well as effective therapeutics for the devastating health burdens which could be based on new technological advancements such as proteomics and nanotechnology.

Proteomics deals with the identification, quantification and characterization of the total protein content present at a given time to help understand life at molecular level by development of novel therapeutic agents and diagnostic tools to provide insights for new biotechnological advancements [14]. The skills, experimental

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approaches and technological platforms supporting proteomics research are rapidly evolving [15]. Therefore, the application of novel qualitative and quantitative nanotechnological findings in the study of NDDs at different levels of neuronal circuit have the potential to help elucidate the biochemical pathogenesis of neurodegeneration and aid in the discovery of new biomarkers [16, 17].

The field of nanotechnology is an amalgamation of chemistry, engineering, biology and medicine [18]. According to the National Cancer Institute (NCI), nanotechnology includes the utilization of various technologies such as nanoarrays, protein arrays, nanopore technology, nanosensors and immunoassays involving nanoparticles (NPs) which have great potential to transform modern medicine in terms of diagnosis and treatment of diseases [19]. Analyses at nanolevel concentrations promise better efficiency, rapidity, low running cost and small sample volume requirement, all of which are indispensible in the latest platforms of proteomics, glycomics and metabolomics [20]. Therefore, the future potential lies in proteomics-based discovery and detection of nano-enabled biomarkers. A rational combination of proteomics with nanotechnology offers greater hope for therapeutic advances that could ameliorate NDDs [21, 22].

Recent advancements in high-throughput genomics and proteomics technologies can aid scientists to develop nano-analytical techniques capable of detecting hormones, drug residues, RNA, DNA, antibodies, peptides, proteins, pathogens and xenobiotics at nanolevel concentrations [23]. New methods and techniques such as protein conjugation with wheat germ agglutinin, cationic moieties (cationization), antibodies and nanogels are being developed for targeted-delivery of drugs and biomacromolecules to the central nervous system (CNS) for treatment of NDDs [24, 25]. Since the use of a number of approaches in understanding the pathogenesis and management of NDDs has not yielded promising results so far, this review focuses on the potential usage of modern proteomics

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and nanotechnological approaches towards a better understanding and management of NDDs. A brief outline of the metabolism and disposition of nano-drugs and nano-enabled drug delivery system is also presented.

2. PROTEOMICS ASPECT OF NDDs

NDDs are pathologically characterized by the progressive formation of lesions composed of disease-specific misfolded proteins [4, 26]. Antemortem prediction of NDDs' pathology is often challenging due to overlapping features of their clinical syndromes [27]. AD is an age-related NDD which is histopathologicallycharacterized by the presence of neurofibrillary tangles (NFTs), senile plaques (SP) and loss of synapse [25]. Over the years, AD has been reported to be one of the most prominent NDD affecting almost 28 million people worldwide [28]. AD is a progressive brain disorder characterized by an irreversible loss of neurons and diminished intellectual abilities such as memory and reasoning thus hampering social or occupational functioning of affected individuals [1]. This chronic illness progresses rather slowly for many years and can manifest in a variety of neurological and psychiatric disorders [29, 30]. The amyloid cascade hypothesis has dominated the field of AD for many years with modern approaches to counter the disease often ending in failures [31]. Over the years, many other hypotheses have also been proposed for the pathogenesis of AD, but none can solely account for overall dimension of this deforming disease [32, 33]. Hence, the exact mechanism of AD pathogenesis still remains elusive.

AD is characterized by certain molecular signals which can only be successfully diagnosed during post-mortem thus further impeding the early diagnosis [34]. Amyloid plaques and NFTs have been implicated as possible neurodegenerative agents of AD [35]. In addition, AD patients are also characterized by a decrease in the brain volume [36]. The main proteinaceous constituent of amyloid plaques is A β , which is 40–42 residue long peptide derived from amyloid precursor protein (APP) [37]. It is believed that APP is deposited at nerve terminals with some studies indicating its role in axonal trafficking [38, 39].

PD is another very prominent and debilitating NDD, which is also characterized by the presence of neuronal loss [40-44]. In PD, degeneration of the midbrain nigrostriatal dopaminergic neurons occurs, thus affecting several important motor symptoms leading to rigidity, bradykinesia, hypokinesia and a resting tremor [45]. Like AD, the exact cause of PD is also not well established. Many factors such as reactive oxygen species (ROS) formation, neuroinflammation and protein misfolding have been implicated in the development of PD [46]. For example, the presence of Lewy bodies, sporadicity and loss of dopaminergic neurons in the substantia nigra are the major characteristics of PD [47]. Owing to the significant research work conducted in the past few decades, the underlying mechanisms of PD are attributed to the right identification of a number of gene-encoding proteins such as a-synuclein, parkin, ubiquitin carboxy-terminal hydrolase L1 (UCH-L1), PARK6, PTEN-induced kinase 1 (PINK1) and DJ-1 [48-53]. These genes have been implicated in protein misfolding, development of oxidative stress and impairment of the ubiquitin-proteasome system [48-55]. In addition, inherited genetic mutations and the corresponding proteins α-synuclein [56], DJ-1 [57], parkin and PINK1 [58] have also been reported to be involved in the pathology of PD.

Many NDDs, especially the PD, are characterized by the presence of α -synuclein clumps in nerve cells. In some familial forms of PD, mutation in the α -synuclein gene [56, 59] and accumulation of α -synuclein protein in the brain have also been reported [60]. For example, a German family with PD was reported to exhibit a missense mutation, Ile93Met, in the *UCHL1* gene [61]. This mutation slightly affects the catalytic activity of the thiol protease and may disturb the normal proteolytic pathway and protein aggregation [62-64] thus suggesting that abnormal aggregation of α -synuclein protein may also be a potential causative factor for PD. Mutations in another protein (*PINK1*) have been reported to be responsible for hereditary early-onset PD [58]. Two homozygous mutations in the putative serine/threonine kinase domain of the PINK1 gene were identified by sequence analysis of candidate genes among PD patients [58]. It was postulated that these mutations may affect substrate recognition or kinase activity thus laying the foundation for the onset of PD [58].

In addition to the above, the pathogenetic mechanisms of many NDDs have been attributed to altered phosphorylation [65, 66]. For example, reports of α -synuclein phosphorylation (serine 129) in the Lewy bodies in human brains with synucleinopathy further suggests the potential role of altered phosphorylation in the pathogenesis of PD [67]. It is postulated that PINK1 may phosphorylate mitochondrial proteins in response to cellular stress in order to confer protection against mitochondrial dysfunction [52, 58, 68]. It has also been reported that, PINK1 is upregulated in cancer cells due to the presence of tumor suppressor gene PTEN [69]. In the neurons, PTEN signaling pathway is reported to promote excitotoxin-induced apoptosis in the hippocampus by regulating cell cycle and cell migration [70]. However, PINK1 was not reported to show any significant effects on PTEN-dependent cell phenotypes [69] thus calling for further investigation on its actual contributory role in the PTEN pathway. The proteins involved in the pathogenesis of AD and PD are summarized in (Fig. 1).

3. PROTEOMIC TOOLS IN NANOTECHNOLOGY

Over the years, many proteomic tools have been modified according to their usage in nanotechnological methods with the most prevalent being the nano-chromatography and nano-electrophoresis (Fig. 2).

3.1. Nano-Chromatography

Nano-chromatography is a combination of chromatographic and capillary electrophoretic separation methods showing high sensitivity (detection of at least up to ng/L). Two types of nano-chromatographic methods are discussed below.

3.1.1. Nano Liquid Chromatography (NLC)

The concept of NLC was introduced by Karlsson and Novotny in 1988. NLC is defined as the chromatographic modality involving samples in nanolitres with detection at the level of ng/ml [71]. NLC is usually performed on a chip and is also known as "lab-on-chip" chromatography. Tubular or packed capillaries with mass spectrometer detector are used to obtain separations in nano-level concentrations. The internal diameter size of the NLC columns are usually 50-100 μ m and offer a good separation potential, hence, they are strategically coupled to micro- and nano-electrospray mass spectrometry [72-74].

Siviero *et al* presented an innovative, reliable and completely automated approach for generation of NLC gradient where the system is electronically-controlled with multi-position valve hosting six loops, each filled with several different mobile phase compositions [75]. It has a low flow rate and valve actuation facility, reducing solvent consumption by 40 times, thus making it attractive since it is economical and environment-friendly as well.

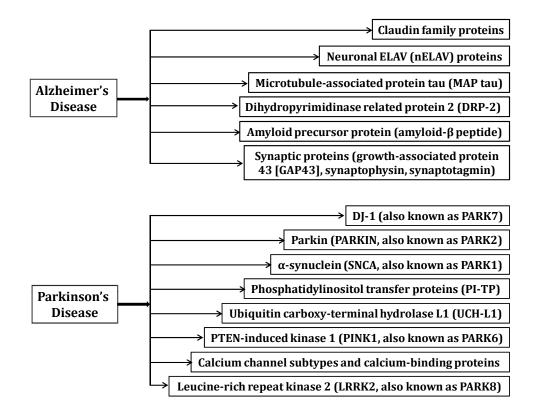


Fig. (1). Proteins involved in the pathogenesis of AD and PD.

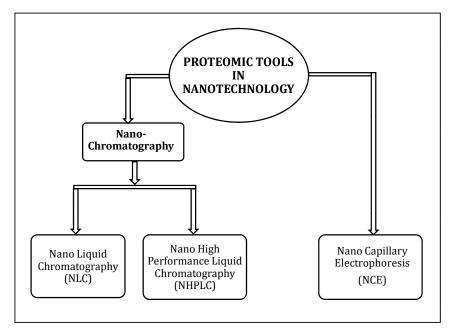


Fig. (2). Various proteomic tools that can be applied in nanotechnology.

3.1.2. Nano High Performance Liquid Chromatography (NHPLC)

A chip-based analytical system which is capable of bearing pressures of between 13 and 150 bars with pressure- or voltagedriven nanochromatography has been developed by Szekely and Freitag allowing microflow sensing and calibration compensation, thus ensuring consistent performance [76]. The mobile phase reservoirs applied in NHPLC are small, air-tight, contaminant free containers made of high quality glass. The mobile phase flows in a constant laminar fashion and usually consists of water/acetone or methanol mixtures. The reproducibility and accuracy of its flow rate are precisely controlled by a microchip flow sensor which is characterized by a high precision, digital intelligence and excellent reliability (Dionex Corporation, USA).

NHPLC employs high grade polyether ether ketone tubings having diameters in micrometer range with minimum connection gaps that can provide a constant laminar flow and minimal void volume. The flow rate of NHPLC pumps ranges between 25 to 4,000

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nL/min. A microchip-based nano-flow sensor has also been designed using a nano-flow splitting technique to generate a slow flow (Dionex Corporation, USA) [77]. In another design, the pressure of compressed air and the flow sensor have been employed for precise nL/min flow rates thereby ensuring an excellent feedback control system (Eksigent, California, USA) [78]. Alternatively, a low dispersion injection method and micro-autosampler can further yield sharp peaks with improved chromatographic resolutions. Overall, the selection of column inner diameter and length and chip is dependent on its applications. Nevertheless, on-chip pressuredriven NHPLC technology still requires further refinement to combine various parts of NHPLC system, especially the separation column, continuous monolithic bed and NP-coated column [79-81] for better efficiencies.

In another study, HPLC coupled to electrochemical detection and nano electro-spray ionization double quadrupole orthogonal acceleration time of flight mass spectrometry were used for identification and quantification of plasma levels of endogenous morphine and anti-inflammatory cytokine, i.e., interleukin-10 as well as adrenocorticotropin levels in PD patients [82]. The study demonstrated that enhanced motor skills and mood elevation is seen with cyclical exercises leading to alleviation of some of the clinical characteristics of PD.

3.2. Nano Capillary Electrophoresis (NCE)

Proteomic analysis especially of low abundant proteins is tedious and exigent. Fortunately, the recent advent of innovative microfluidic devices such as NCE can help ease the challenge [83, 84]. Protein digestion by enzymatic treatment plays an important role in the sample pre-treatment process. Usually, proteins are fragmented into small peptides prior to analysis. The chip-based capillary electrophoresis (CE) involve fused silica capillaries thus allowing nanolevel analyses of low quantity samples or samples present in minimal concentrations for genomic, proteomic and drug development studies. It comprises a microchannel network for pre- and postsample handling, reactions, separation and identification. NCE is characterized by sample injection and electrolyte flowing at nanolevel rate thereby reducing sample volume and imparting high speed and improved separation efficiencies [85, 86].

A new portable microchip electrophoresis, equipped with a high voltage power supply having dual amperometric (DC or pulsed) detection capability, a bipotentiostat and a chip holder has been designed for *in situ* analysis using microchips signal transduction by electrochemical detection [87]. Its performance was reported to be better than other commercial gadgets for separation of neuro-transmitters, epinephrine, 3,4-dihydroxy-L-phenyl-alanine and dopamine.

A two chip-based NCE system was microfabricated by Phillips *et al* for analyses of inflammatory neuropeptides in body fluids [88]. Dual chips are designed to perform electrokinetic flow immunoassays by utilizing an immunoaffinity port containing an array of immobilized antibodies to rapidly and accurately capture the analytes of interest (neuro-inflammatory biomarkers in complex biological fluids). It is postulated that with the ever increasing array of commercially-available antibodies, the chip-based system may be in the diagnosis of various NDDs.

Insulin degrading enzyme (IDE) is the main enzyme responsible for A β clearance from the brain [89]. IDE-mediated A β proteolysis is a progressive enzymatic process subjected to alternative substrate inhibition, especially by insulin. In another study, a CE method for *in vitro* investigation of IDE-mediated A β_{1-40} proteoly-

sis employing only a conventional CE instrument equipped with a fused silica capillary has also been reported [90]. It is hypothesized that further developments in this technique will be of major significance to biomedical utility in the future.

4. PROTEOMIC APPROACHES IN THE MANAGEMENT OF NDDs

High-throughput proteomics approaches are utilized to elucidate new therapeutic biomarkers which can be potential drug targets. While designing effective therapeutics for NDDs, rapid qualitative and quantitative analysis of neurotransmitters is a major concern. Vlc'kova' and Schwarz developed a quick but sensitive (3-8 times higher sensitivity) separation and detection method for catecholamines having similar structures such as noradrenaline, dopamine, adrenaline and their O-methoxylated metabolites including 3-methoxytyramine, normetanephrine and metanephrine from the mouse brain homogenate using a complex phosphate-borate buffer with sodium dodecyl sulfate (SDS) and polyamidoamine dendrimer and by using CNT-film-modified gold electrode detector [91]. Overall, carbon nanotube-based detectors have been applied in microchip and CE systems for the detection of neurotransmitters by several group of researchers [91-93].

In addition, the presence of $A\beta_{(1-40)}$ and $A\beta_{(1-42)}$ in the cerebrospinal fluid and plasma have been proposed as potential biomarkers of AD [94]. Therefore, quantification of circulating A β peptides in plasma can facilitate AD diagnosis. To determine the most suitable technique for detecting A β_{1-40} levels in plasma or serum, Varesio *et al* compared NLC and CE attached to mass spectrometers [95]. A 50 µm I.D. CE capillary and a 75 µm I.D. NLC column were coupled to a single quadrupole mass spectrometer with a sheath-liquid electrospray interface and a nanospray interface, respectively [95]. It was concluded that CE has comparatively lower sensitivity, thus limiting its usage in biological matrix analyses. On the other hand, NLC has column switching set-up and higher sample loading capability, thus allowing better sensitivity of detection of $A\beta_{1-40}$ when used at ng/ml concentrations [95].

For effective treatment of NDDs, the levels of fluid $A\beta_{1-42}$, tau and phosphorylated tau can be good indicators [96]. Blood-brain barrier (BBB) acts as the biggest hurdle in the delivery of therapeutic proteins to the CNS [97]. Brain microvessel endothelial cells (BMVEC) which forms tight extracellular junctions and have low pinocytic activity are mainly responsible for the low permeability seen with the BBB [24]. Therefore, the development of new innovative methods and techniques for effective delivery of drugs and bio-macromolecules to the CNS is the primary requirement for the treatment of NDDs [98].

A plethora of methods and techniques are also applied in protein modifications so as to enhance penetration into the BBB. For this purpose, proteins have been conjugated with wheat germ agglutinin [99, 100], cationic compounds [101-103] and transferrin receptors [104-107]. In addition, antibodies were also conjugated with insulin for similar function. Even though the above mentioned strategies are rather successful in increasing the uptake of proteins into the CNS via adsorptive endocytosis, various aspects such as toxicity and antigenicity of such modified proteins can pose as limiting factors [104]. In cationization process, superficial carboxyl groups present in the protein is converted into an extended primary amino group which can be used to enhance the interaction of modified proteins with negative charge substances at the luminal plasma membrane of the brain endothelial cells. The cationized protein then undergoes adsorptive transcytosis occurring via the BBB. Cationization of antibodies can be achieved by using several synthetic (hexamethylenediamine) or naturally occurring (e.g., putrescine) polyamines with the later being the most efficient. Nevertheless, although this approach is not free from some drawbacks, it is still a realistic approach for transportation of antibodies across the BBB [101-103]. In another successful example of conjugation, the OX26 antibody was conjugated with basic fibroblast growth factor and brain-derived neurotrophic factor and showed promising results in cerebral trauma models [108-110].

Protein transduction domain (PTD) is another significant approach for cellular delivery of polypeptides, polynucleotides and NPs. PTDs are small cationic peptides which can facilitate the uptake of large, biologically-active molecules into mammalian cells. They are significant because they can eliminate the problems posed by size restriction shown by some useful but otherwise larger drug molecules, thus enabling previously unavailable drugs to modulate and alleviate several diseases [111, 112]. Several sources of PTDs have also been explored including human immunodeficiency virus-1-transcriptional activator (HIV-1-TAT) peptide, DNA-binding protein (VP22), Drosophila Antennapedia (Antp) homeotic transcription factor and herpes simplex virus-1 (HSV-1). The HIV-1-TAT peptide is a small basic peptide successfully shown to deliver a large variety of molecules including small particles, proteins, peptides and nucleic acids. However, the region displaying properties of good cell penetration appears to be confined only to a small stretch (RKKRRQRRR) of 9 basic amino acids [113]. PTDs can be used in several ways. They can be introduced into protein by chemical conjugation method or alternatively, can be geneticallyfused to the protein cDNA and expressed in host mammalian cells via transfection or they can also be produced in bacteria even though the mechanism of transduction still remains unknown [112, 114, 115]. To date, various TAT fusion proteins have been investigated for the treatment of NDDs [116]. Some successful examples include the intravenous delivery of TAT peptide conjugated with antiapoptotic factor Bcl-XL and glial cells line derived neurotropic factor (GDNF), TAT-GDNF fusion protein in MPTP (1-methyl-4phenyl-1,2,3,6- tetrahydropyridine) model of AD [117-119]. Another PTD fusion protein conjugated with tyrosine hydroxylase showed significant effects on 6-hydroxydopamine induced PD model rats [120]. In other words, PTD conjugated protein modification strategy improves CNS drug delivery significantly, but its potential as treatment for NDDs is limited by its immunogenicity and long term side effects.

When compared to smaller molecules, the transport of biomacromolecules such as DNA or proteins across the BBB is more challenging. Nevertheless, naturally-occurring peptides which have the ability to effectively pass this barrier by receptor-mediated endocytosis (RME) provide some hope in this regard [121, 122]. The specific peptides can be used for targeting biomolecules by RME and can be selected from a phage display library [123]. Conjugation of these peptides with an appropriate drug carrier molecule further facilitate its transport [124]. However, limitation in size not exceeding 100 nm is the primary requirement for carrier molecules. Besides the above, NPs can effectively be used as a vehicle for drug and gene deliveries [125-127]. Galectins can also be conjugated with NPs as they have been reported to have significant roles in NDDs by virtue of their wide spectrum of properties [128-130]. Other examples include solid NPs [126, 131, 132], liposomes [133, 134] and polymer micelles [135-138]. The modification of carrier by polyethylene glycol (PEG) can be carried out to improve the stability of NPs in dispersion and increase their bioavailability

[139-142]. Peptides and proteins can be attached to the end of PEG chains to facilitate receptor-mediated bindings. In addition, antibody and insulin-conjugated micelles have been found to be very effective in in vivo drug delivery to the brain tissue [143]. Immunoliposomes carrying therapeutic antisense epidermal growth factor receptor (EGFR) gene have been reported to deliver substances to EGFR-dependent brain gliomas in vivo successfully [144]. PEGylated immune-liposomes containing antibodies directed to insulin or transferrin receptors have been successful as carriers for gene replacement in PD model [145]. The PEGylated immune-liposomes are also used for targeting and transfecting β -galactosidase (LacZ) and luciferase into the brain [146, 147]. In another exciting strategy, a new family of carrier system called nanogel has been introduced for specific targeting of drugs and biomolecules to the brain [148]. Nanogels are cross-linked polymers often made by a combination of ionic and non-ionic polymer chains and are prepared following emulsification using a solvent evaporation system [149, 150]. Nanogels tend to swell in the presence of water and can incorporate several biomolecules including the oligos, siRNA, proteins, DNA and drug molecules with an encapsulation efficiency of approximately 40-60%, and they also have the ability to decrease possible degradation of biomolecules occurring during transportation [151]. Nanogels can also interact with a large number of biomolecules such as negatively-charged peptides and proteins. The surface of nanogels can be modified in many ways by using either transferrin or insulin by avidin-biotin coupling or for targeting receptors present at BMVEC.

For the treatment of NDDs such as AD and PD [152, 153], lysosomal diseases [154, 155] as well as obesity [121, 156], there is an urgent need to augment delivery of therapeutic peptide and proteins to the brain, besides using them as targeted moieties [152, 153]. RME (such as insulin, insulin-like growth factor and transferrin) is utilized by known peptides in order to cross the BBB to a remarkable extent [157]. For example, in artificial hydrophobization, a very small number of fatty acid residues (stearic, palmitic and oleic acid) can be conjugated with protein molecules [158-161]. In this process, a protein molecule is altered in a system of reversed micelles due to the presence of water-insoluble reagent such as fatty acid chloride. Fatty-acylated proteins then acquire the ability to translocate across plasma membranes and penetrate into intact cells. Protein conjugation with controlled modification can be achieved by using reversed micelles, resulting in an increased binding of these proteins with the lipid membranes as a result of anchoring of the hydrophobic groups [157]. With this approach, the protein molecule acquires hydrophobic anchor groups which can specifically target hydrophilic proteins to the cell surfaces besides remaining to be water soluble. By utilizing this technique, more than a dozen of proteins have been modified with their functional activities remaining intact [161-168]. Insulin modified with a palmitic acid residue is a successful example of enhanced hypoglycemic effect generation since it was less immunoreactive when compared to the native insulin [160]. Chekhonin et al reported that the interaction between fatty acid residue and antigen binding site is vital for the delivery of modified antibodies to the brain [165, 169]. A group of researchers in France synthesized a fatty acylated ribonuclease A (Rnase A) with the ability to cross BMVEC monolayer with minor degradation [170]. Furthermore, stearoylation of Rnase A [170] or horseradish peroxidase [171] significantly increases their penetration across the BBB. Amongst the fatty acid derivatives, stearic acid was found to be most active [170]. Protein conjugation with Pluronic® block copolymers is another successful strat-

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egy/approach to successful delivery of drugs and biomolecules across the CNS [171]. The block copolymers are composed of hydrophilic ethylene oxide and hydrophobic propylene oxide arranged in a particular pattern which can thus interact simultaneously with hydrophobic surfaces as well as the plasma membrane due to their amphipathic nature. Their mode of action includes inhibition of drug efflux transporters expressed in BBB, thus allowing increased transport of various substances to the brain [172, 173]. Overall, the above approaches are very promising for improving drug delivery of protein-based diagnostic and therapeutic agents to the CNS. A schematic representation of nanotechnology-based drug delivery approaches in the treatment of NDDs is depicted in (Fig. **3**).

5. NANOTECHNOLOGICAL APPROACHES IN THE MAN-AGEMENT OF NDDs

The mysteries associated with various NDDs may be unraveled by newly emerging nanotechnological approaches, since some of the methods have been reported to correlate with NP exposures. A positive result in this aspect shows that the damaging effect of human exposure to toxic NPs can be reduced by identifying creationexposure pathways of toxins [174]. On the other hand, the toxic properties of some NPs may allow treatment of diseases at cellular level and could potentially be utilized for the treatment of various NDDs.

Among currently available therapies for various NDDs, the oral administration of dopamine agonists such as levodopa is the most common. However, the effects of levodopa along with the motor side-effects tend to decline among NDD patients with disease progression. Therefore, transplantation of fetal dopamine neurons and deep brain stimulation has been explored to complement the pharmacological treatments [175, 176]. Nevertheless, these approaches still remain inconclusive and are not very effective in halting the continual loss of dopamine neurons [177, 178] thus creating the need of new approaches to combat the progression of NDDs.

Nanotechnological approach can be a potential way towards further understanding and management of various NDDs. In this regards, a number of nanostructures have been employed for the development of nano-enabled drug delivery. Some of the nanostructures already in use include polymeric Nps [179], polymeric nanospheres and nano-suspensions [180], polymeric nano-gels [150], carbon nano-tubes and nano-fibers [44], polymeric nano-micelles [181] and as well as polymeric nano-liposomes [44]. Even though these approaches have shown some promising results, more concrete scientific efforts remain desired. Table **1** lists the various types of nanostructures used in the treatment of NDDs.

6. METABOLISM AND DISPOSITION OF NANO-DRUGS

Nano-drugs are the result of the interplay between nanotechnology and modern medicine (nanomedicine). Nano-drugs adsorb NPs on their surface and are specifically targeted to a particular cell or organ to provide maximum safety with minimal side effects [182-184]. Nano-drugs have the advantage of overcoming the natural barriers present in our body defense system which generally can pose several challenges to drug delivery. Nano-drugs can enter the capillaries, penetrate cells, get absorbed though pinocytosis and can enhance bioavailability [97]. Due to their large surface area, nanodrugs can further enhance the solubility of poorly soluble drugs and also increase their half-lives by controlling the speed of degradation in vivo, thus augmenting drugs' efficacy and lowering their side effects [184-187]. After entering the body, drug and NPs get separated under a constant speed thus creating a time lapse before they reach their targets [188]. Nano-drugs have been reported to reach into specific body parts mainly by infiltration, leaching and proliferation (dissolution) [189]. The conjugation of drugs with NPs prevents enzymatic degradation of drugs, increases effective drug release time, reduces side effects and improves efficiency.

Some of the effective methods of controlled nano-drugs release include chemical, solvent and diffusion control [190]. Among these, diffusion control is the most commonly used method for nondegradable polymeric carriers [191-193]. For the chemical control method, hydrolysis and other types of chemical reactions efficiently reduces time and rate of drug release [194]. Depending upon the role of drugs and NPs involved, chemical control method utilizes two different approaches; the side-chain and the degradable systems [97]. The rate of polymer degradation by different enzymes is the limiting factor for nano-drug release rate in the degradable system [195]. Following degradation, the metabolites can be absorbed or discharged by the body, thus regulating polymer degradation at a particular location within a regular time frame [196]. The limiting factor is the polymer degradation rate which not only affects the release of nano-drugs in a specified time, but is also

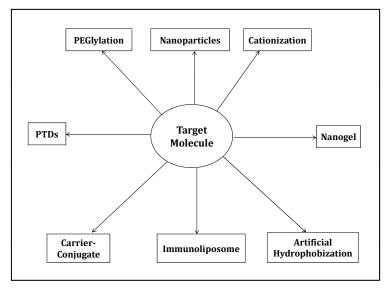


Fig. (3). Schematic representation of nanotechnology-based drug delivery approaches in the treatment of various NPs.

Table 1. Nanostructures used in treatment of neurodegenerative disorders. (Aβ, amyloid-β; AD, Alzheimer's Disease; Ach, Acetylcholine; NGF, Nerve Growth Factor; PD, Parkinson's Disease).

Nanostructure	Target	Drug Delivered	References
Nanocapsules & nanospheres	Neuroinflammation	Indomethacin	[209, 210]
	Aβ in AD	Thioflavin-T	[211]
	Aβ in AD	Clioquinol	[212]
	Aβ in AD	d-Penicillamine	[213]
Nanogels	Brain delivery	Oligonucleotides	[149, 151]
	Aβ in AD	Cholesterol bearing pullunan	[209]
Carbon nanotubes & nanofibers	Nerve growth	NGF	[214]
	Dopamine in PD	Biosensor	[214]
	AD	Ach	[215]
Nanomicelles	Aβ in AD	PEGylated phospholipids	[216]
Nanoliposomes	$A\beta$ in AD	Curcumin	[217]

dependent on other factors such as quality of the polymer, its molecular weight, crystalline nature of NPs, hydrophilicity and hydrophobicity [197]. In the side-chain release method, both degradable and nondegradable types of nano-drug carrier can be used. The side chain which is attached via some chemical bonds can be broken by hydrolysis or enzymes, thereby controlling the release of drugs [197]. Beside these factors, the release of nano-drugs is affected by several other parameters, such as the nature and composition of polymers and nano-drugs, surrounding temperature and environment, pH, and hydrophilicity and hydrophobicity of degradable polymers.

A major concern with the use of nano-drugs is the evaluation of their biosafety and toxicity [198-202] profiles. To date, there are only few studies related to the toxicity of NPs that are in use [203-205]. Overall pharmacodynamics, tissue distribution, plasma clearance and urinary excretion of nano-drugs still need some careful evaluation [206]. Nanomaterials consist of metal components like quantum dots, nano-gold, nano-silver and nano-zinc oxide. When these metal-based nanomaterials are used for biological applications, their biosafety must be monitored. Furthermore, the biological disposition including the ADMET (absorption, deposition, metabolism, elimination and toxicity) of the nanomaterials needs to be further evaluated. Such evaluation can be done by tracking the changes in the metallic constituents of NPs in various tissues and organs following exposure. Atomic absorption spectrometry (AAS) and inductively-coupled plasma mass spectrometry (ICP-MS) are the preferred techniques for metal analyses [207].

Four mechanisms of NP toxicity have been identified: 1) toxicity of any constituents present 2) toxicity of their degradation products 3) toxicity as a result of endocytosis of NPs and 4) toxicitymediated membrane lysis [196]. The U.S. Food and Drug Administration (FDA) have developed a new task force on nanotechnology, but more insight or regulation needs to be practiced. Researchers from all over the world have expressed their anxiety about the harmful effects of nano-drugs on human health [196]. This is due to the fact that the minute size of NPs theoretically can allow their infiltration into all body cells and can therefore be potentially detrimental to healthy cells. Another vital concern is the proper disposal of NPs used in the manufacturing or other processes. Special disposal techniques are required to avoid harmful particles from accumulating in the environment following which monitoring may be an uphill task if not impossible [208].

7. CONCLUSION

The limited available therapeutic options for NDDs have extended some new therapeutic prospects. Modern nanotechnological approaches coupled with new proteomic tools promise to be a revolutionizing way of further understanding and managing these disorders. With the recent advent of carbon nanotube-based detectors, rapid qualitative and quantitative analyses of neurotransmitters are feasible. One good example is the excellent NP-based system as drug (nanodrugs) and gene delivery vehicles to penetrate the unique BBB layer. These are rather promising in exploring novel approaches in early protein-based diagnostics focusing on the detection of preinflammatory states and plaque characterization as well as new therapeutic agents. Reducing toxicity and increasing target specificity of nano-drugs will hopefully transform numerous facets of brain physiological studies and clinical neurology in the future.

CONFLICT OF INTEREST

The authors confirm that this article content has no conflict of interest.

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LIST OF ABBREVIATIONS

NDD	=	Neurodegenerative disorder
AD	=	Alzheimer's disease
PD	=	Parkinson's disease

Αβ	=	Amyloid-β
BBB	=	Blood-brain barrier
CNS	=	Central nervous system
NFTs	=	Neurofibrillary tangles
APP	=	Amyloid precursor protein
PINK1	=	PTEN-induced kinase
UCH-L1	=	Ubiquitin carboxy-terminal hydrolase L1
IDE	=	Insulin degrading enzyme
NPs	=	Nanoparticles
NLC	=	Nano liquid chromatography
NHPLC	=	Nano high performance liquid chromatography
NCE	=	Nano capillary electrophoresis
CE	=	Capillary electrophoresis
PTD	=	Protein transduction domain
PEG	=	Polyethylene glycol
BMVEC	=	Brain microvessel endothelial cells
RME	=	Receptor-mediated endocytosis

REFERENCES

- Mirza, Z.; Ali, A.; Ashraf, G.M.; Kamal, M.A.; Abuzenadah, A.M.; Choudhary, A.G.; Damanhouri, G.A.; Sheikh, I.A. Proteomics approaches to understand linkage between Alzheimer's disease and type 2 diabetes mellitus. *CNS Neurol. Disord. Drug Targets*, 2014, *13*(2), 213-225.
- [2] Kanazawa, I. How do neurons die in neurodegenerative diseases? Trends Mol. Med., 2001, 7(8), 339-344.
- [3] Lee, S.J.; Lim, H.S.; Masliah, E.; Lee, H.J. Protein aggregate spreading in neurodegenerative diseases: problems and perspectives. *Neurosci. Res.*, 2011, 70(4), 339-348.
- [4] Ashraf, G.M.; Greig, N.H.; Khan, T.A.; Hassan, I.; Tabrez, S.; Shakil, S.; Sheikh, I.A.; Zaidi, S.K.; Akram, M.; Jabir, N.R.; Firoz, C.K.; Naeem, A.; Alhazza, I.M.; Damanhouri, G.A.; Kamal, M.A. Protein misfolding and aggregation in Alzheimer's disease and type 2 diabetes mellitus. CNS Neurol. Disord. Drug Targets, 2014, 13(7), 1280-1293.
- [5] Banu, S.; Jabir, N.R.; Manjunath, N.C.; Khan, M.S.; Ashraf, G.M.; Kamal, M.A.; Tabrez, S. Reduction of post-prandial hyperglycemia by mulberry tea in type-2 diabetes patients. *Saudi J. Biol. Sci.*, 2014, (article in press).
- [6] Jabir, N.R.; Firoz, C.K.; Baeesa, S.S.; Ashraf, G.M.; Akhtar, S.; Kamal, W.; Kamal, M.A.; Tabrez, S. Synopsis on the Linkage of Alzheimer's and Parkinson's Disease with Chronic Diseases. CNS Neurosci. Ther., 2014 (article in press).
- [7] Khan, N.M.; Ahmad, A.; Kamal, M.A.; Mushtaq, G.; Ashraf, G.M. Current Challenges to Overcome the Management of Type 2 Diabetes Mellitus and Associated Neurological Disorders. *CNS Neurol. Disord. Drug Targets*, 2014, (article in press).
- [8] Aliev, G.; Burzynski, G.; Ashraf, G.M.; Jabir, N.R.; Cacabelos, R.; Benberin, V.V.; Burzynski, S.R. Implication of oxidative stressinduced oncogenic signaling pathways as a treatment strategy for neurodegeneration and cancer. In *Sys. Bio. Free Rad. Antioxid.* Laher, I., Ed.; Springer Berlin Heidelberg, **2014**, pp 2325-2347.
- [9] Adnan Ahmad, S.S.; Ghulam M. Ashraf, S.T. A Region-specific Treatment Strategy To Address The Problem Of Drug Resistance By NDM-1-producing Pathogens. *Enz. Eng.*, 2013, 02(01), 1-3.
- [10] Ali, R.; Mirza, Z.; Ashraf, G.M.D.; Kamal, M.A.; Ansari, S.A.; Damanhouri, G.A.; Abuzenadah, A.M.; Chaudhary, A.G.; Sheikh, I.A. New anticancer agents: recent developments in tumor therapy. *Anticancer Res.*, **2012**, *32*(7), 2999-3005.
- [11] Hasan, S.S.; Ashraf, G.M.; Banu, N. Galectins Potential targets for cancer therapy. *Cancer Lett.*, 2007, 253(1), 25-33.
- [12] Sharma, A.; Kumar, R.; Varadwaj, P.K.; Ahmad, A.; Ashraf, G.M. A comparative study of support vector machine, artificial neural network and bayesian classifier for mutagenicity prediction. *Interdisc. Sci., Comp. Life Sci.*, 2011, 3(3), 232-239.

- [13] Rajnish Kumar, A.S. Classification of oral bioavailability of drugs by machine learning approaches: a comparative study. J. Comp. Interdisc. Sci., 2011, 2(9), 1-18.
- [14] Chandramouli, K.; Qian, P.Y. Proteomics: Challenges, Techniques and Possibilities to Overcome Biological Sample Complexity. *Hum. Genomics Proteomics*, 2009, 1-22.
- [15] Zhang, J.; Keene, C.D.; Pan, C.; Montine, K.S.; Montine, T.J. Proteomics of human neurodegenerative diseases. J. Neuropathol. Exp. Neurol., 2008, 67(10), 923-932.
- [16] Kroksveen, A.C.; Opsahl, J.A.; Aye, T.T.; Ulvik, R.J.; Berven, F.S. Proteomics of human cerebrospinal fluid: discovery and verification of biomarker candidates in neurodegenerative diseases using quantitative proteomics. *J. Proteomics*, **2011**, *74*(4), 371-388.
- [17] Ramos-Cabrer, P.; Campos, F. Liposomes and nanotechnology in drug development: focus on neurological targets. *Int. J. Nanomedicine*, 2013, 8, 951-960.
- [18] Jabir, N.R.; Tabrez, S.; Ashraf, G.M.; Shakil, S.; Damanhouri, G.A.; Kamal, M.A. Nanotechnology-based approaches in anticancer research. *Int. J. Nanomedicine*, **2012**, *7*, 4391-4408.
- [19] Tabrez, S.; Priyadarshini, M.; Urooj, M.; Shakil, S.; Ashraf, G.M.; Khan, M.S.; Kamal, M.A.; Alam, Q.; Jabir, N.R.; Abuzenadah, A.M.; Chaudhary, A.G.A.; Damanhouri, G.A. Cancer chemoprevention by polyphenols and their potential application as nanomedicine. J. Environ. Sci. Health C Environ. Carcinog. Ecotoxicol. Rev., 2013, 31(1), 67-98.
- [20] Novotny, M.V.; Soini, H.A.; Mechref, Y. Biochemical individuality reflected in chromatographic, electrophoretic and massspectrometric profiles. J. Chromatogr. B Analyt. Technol. Biomed. Life. Sci., 2008, 866(1-2), 26-47.
- [21] Rakowska, P.D.; Ryadnov, M.G. Nano-enabled biomarker discovery and detection. *Biomark. Med.*, 2011, 5(3), 387-396.
- [22] Jain, K.K. The role of nanobiotechnology in drug discovery. Drug Discov. Today, 2005, 10(21), 1435-1442.
- [23] Nanochromatography and Nanocapillary Electrophoresis: Pharmaceutical and Environmental Analyses (http://onlinelibrary.wiley. com/book/10.1002/9780470434925).
- [24] Mayhan, W.G. Regulation of blood-brain barrier permeability. *Microcirculation (New York, N.Y.: 1994)*, 2001, 8(2), 89-104.
- [25] Selkoe, D.J. Alzheimer's disease: genes, proteins, and therapy. *Physiol. Rev.*, 2001, 81(2), 741-766.
- [26] Serrano-Pozo, A.; Frosch, M.P.; Masliah, E.; Hyman, B.T. Neuropathological Alterations in Alzheimer Disease. *Cold Spring. Harb. Perspect. Med.*, 2011, 1(1), 1-23.
- [27] Hu, W.T.; Chen-Plotkin, A.; Arnold, S.E.; Grossman, M.; Clark, C.M.; Shaw, L.M.; Pickering, E.; Kuhn, M.; Chen, Y.; McCluskey, L.; Elman, L.; Karlawish, J.; Hurtig, H.I.; Siderowf, A.; Lee, V.M.Y.; Soares, H.; Trojanowski, J.Q. Novel CSF biomarkers for Alzheimer's disease and mild cognitive impairment. *Acta Neuropathol. (Berl.)*, **2010**, *119*(6), 669-678.
- [28] Sheikh, I.A.; Ali, R.; Dar, T.A.; Kamal, M.A. An overview on potential neuroprotective compounds for management of Alzheimer's disease. CNS Neurol. Disord. Drug Targets, 2012, 11(8), 1006-1011.
- [29] Kaminsky, Y.G.; Reddy, V.P.; Ashraf, G.M.; Ahmad, A.; Benberin, V.V.; Kosenko, E.A.; Aliev, G. Age-related defects in erythrocyte 2,3-diphosphoglycerate metabolism in dementia. *Aging Dis.*, 2013, 4(5), 244-255.
- [30] Aliev, G.; Ashraf, G.M.; Kaminsky, Y.G.; Sheikh, I.A.; Sudakov, S.K.; Yakhno, N.N.; Benberin, V.V.; Bachurin, S.O. Implication of the nutritional and nonnutritional factors in the context of preservation of cognitive performance in patients with dementia/depression and Alzheimer disease. *Am. J. Alzheimers Dis. Other Demen.*, 2013, 28(7), 660-670.
- [31] Dar, T.A.; Sheikh, I.A.; Ganie, S.A.; Ali, R.; Singh, L.R.; Gan, S.H.; Kamal, M.A.; Zargar, M.A. Molecular linkages between diabetes and Alzheimer's disease: current scenario and future prospects. CNS Neurol. Disord. Drug Targets, 2014, 13(2), 290-298.
- [32] Butterfield, D.A.; Stadtman, E.R. In Adv. Cell Aging Geron. Paula, S.T.; Bittar, E.E., Eds.; Elsevier, 1997; Vol. Volume 2, pp 161-191.
- [33] Markesbery, W.R. Oxidative stress hypothesis in Alzheimer's disease. Free Radic. Biol. Med., 1997, 23(1), 134-147.
- [34] Jacobs, A.H.; Winkler, A.; Castro, M.G.; Lowenstein, P. Human gene therapy and imaging in neurological diseases. *Eur. J. Nucl. Med. Mol. Imaging*, 2005, 32(Suppl 2), S358-S383.
- [35] Götz, J.; Schild, A.; Hoerndli, F.; Pennanen, L. Amyloid-induced neurofibrillary tangle formation in Alzheimer's disease: insight

from transgenic mouse and tissue-culture models. Int. J. Dev. Neurosci., 2004, 22(7), 453-465.

- [36] Prabakar, S. An investigation of volumetric and corpus callosum dimension to detect brain disorders. J. Biomed. Sci. Eng., 2012, 05(07), 369-377.
- [37] Selkoe, D.J. Alzheimer's disease is a synaptic failure. Science, 2002, 298(5594), 789-791.
- [38] Sabo, S.L.; Ikin, A.F.; Buxbaum, J.D.; Greengard, P. The Alzheimer amyloid precursor protein (APP) and FE65, an APPbinding protein, regulate cell movement. J. Cell Biol., 2001, 153(7), 1403-1414.
- [39] Sabo, S.L.; Ikin, A.F.; Buxbaum, J.D.; Greengard, P. The amyloid precursor protein and its regulatory protein, FE65, in growth cones and synapses *in vitro* and *in vivo*. J. Neurosci., 2003, 23(13), 5407-5415.
- [40] Bensadoun, J.-C.; Pereira de Almeida, L.; Fine, E.G.; Tseng, J.L.; Déglon, N.; Aebischer, P. Comparative study of GDNF delivery systems for the CNS: polymer rods, encapsulated cells, and lentiviral vectors. J. Control. Release, 2003, 87(1-3), 107-115.
- [41] Kishima, H.; Poyot, T.; Bloch, J.; Dauguet, J.; Condé, F.; Dollé, F.; Hinnen, F.; Pralong, W.; Palfi, S.; Déglon, N.; Aebischer, P.; Hantraye, P. Encapsulated GDNF-producing C2C12 cells for Parkinson's disease: a pre-clinical study in chronic MPTP-treated baboons. *Neurobiol. Dis.*, **2004**, *16*(2), 428-439.
- [42] McArthur, J.C. HIV dementia: an evolving disease. J. Neuroimmunol., 2004, 157(1-2), 3-10.
- [43] Menei, P.; Montero-Menei, C.; Venier, M.-C.; Benoit, J.-P. Drug delivery into the brain using poly(lactide-co-glycolide) microspheres. *Expert Opin. Drug Deliv.*, 2005, 2(2), 363-376.
- [44] Kabanov, A.V.; Gendelman, H.E. Nanomedicine in the diagnosis and therapy of neurodegenerative disorders. *Prog. Poly. Sci.*, 2007, 32(8-9), 1054-1082.
- [45] Singh, N.; Pillay, V.; Choonara, Y.E. Advances in the treatment of Parkinson's disease. *Prog. Neurobiol.*, 2007, 81(1), 29-44.
- [46] Kanwar, J.R.; Sun, X.; Punj, V.; Sriramoju, B.; Mohan, R.R.; Zhou, S.-F.; Chauhan, A.; Kanwar, R.K. Nanoparticles in the treatment and diagnosis of neurological disorders: untamed dragon with fire power to heal. *Nanomedicine*, **2012**, *8*(4), 399-414.
- [47] Park, S.S.; Lee, D. Selective loss of dopaminergic neurons and formation of Lewy body-like aggregations in alpha-synuclein transgenic fly neuronal cultures. *Eur. J. Neurosci.*, 2006, 23(11), 2908-2914.
- [48] Dauer, W.; Przedborski, S. Parkinson's disease: mechanisms and models. *Neuron*, 2003, 39(6), 889-909.
- [49] Dawson, T.M.; Dawson, V.L. Molecular Pathways of Neurodegeneration in Parkinson's Disease. *Science*, 2003, 302(5646), 819-822.
- [50] Aliev, G.; Priyadarshini, M.; Reddy, V.P.; Grieg, N.H.; Kaminsky, Y.; Cacabelos, R.; Ashraf, G.M.; Jabir, N.R.; Kamal, M.A.; Nikolenko, V.N.; Zamyatnin, A.A.; Benberin, V.V.; Bachurin, S.O. Oxidative stress mediated mitochondrial and vascular lesions as markers in the pathogenesis of Alzheimer disease. *Curr. Med. Chem.*, 2014, 21(19), 2208-2217.
- [51] Ahmad, A.; Rasheed, N.; Gupta, P.; Singh, S.; Siripurapu, K.B.; Ashraf, G.M.; Kumar, R.; Chand, K.; Maurya, R.; Banu, N.; Al-Sheeha, M.; Palit, G. Novel Ocimumoside A and B as anti-stress agents: modulation of brain monoamines and antioxidant systems in chronic unpredictable stress model in rats. *Phytomed.*, 2012, 19(7), 639-647.
- [52] Ahmad, A.; Rasheed, N.; Gupta, P.; Ashraf, G.M.; Singh, S.; Chand, K.; Maurya, R.; Palit, G. Novel <i>Ocimum sanctum</i>compounds modulate stress response: Role of CRF, POMC, GR and HSP-70 in the hypothalamus and pituitary of rats. *Med. Plants*, 2013, 5(4), 194-201.
- [53] Ahmad, A.; Rasheed, N.; Ashraf, G.M.; Kumar, R.; Banu, N.; Khan, F.; Al-Sheeha, M.; Palit, G. Brain region specific monoamine and oxidative changes during restraint stress. *Can. J. Neurol. Sci. Le*, **2012**, *39*(3), 311-318.
- [54] Aliev, G.; Horecký, J.; Vančová, O.; Ashraf, G.M.; Hassan, I.; Bragin, V.; Bragin, I.; Shevtsova, E.; Klochkov, S.G.; Kosenko, E.A.; Cacabelos, R.; Bachurin, S.O.; Benberin, V.V.; Kaminsky, Y.G. The three-vessel occlusion as a model of vascular dementia – oxidative stress and mitochondrial failure as an indicator of brain hypoperfusion. In Sys. Bio. Free Rad. Antioxid. Laher, I., Ed.; Springer Berlin Heidelberg, 2014, pp 2023-2032.
- [55] Aliev, G.; Ashraf G.M.; Horecký, J.; Vancová, O.; Gvozdjáková, A.; Kucharská, J.; Palacios, H.H.; Li, Y.; Perveen, A.; Khan, T.A.;

Bragin, V.; Bragin, I.; Shevtsova, E.; Klochkov, S.G.; Kosenko, E.A.; Cacabelos, R.; Kaminsky, Y.G.; Sudakov, K.V.; Benberin, V.V.; Bachurin, S.O. Potential Preventive Effects of Coenzyme Q and Creatine Supplementation on Brain Energy Metabolism in Rats Exposed to Chronic Cerebral Hypoperfusion. In *Sys. Bio. Free Rad. Antioxid.* Laher, I., Ed.; Springer Berlin Heidelberg, **2014**, pp 2033-2048.

- [56] Polymeropoulos, M.H.; Lavedan, C.; Leroy, E.; Ide, S.E.; Dehejia, A.; Dutra, A.; Pike, B.; Root, H.; Rubenstein, J.; Boyer, R.; Stenroos, E.S.; Chandrasekharappa, S.; Athanassiadou, A.; Papapetropoulos, T.; Johnson, W.G.; Lazzarini, A.M.; Duvoisin, R.C.; Di lorio, G.; Golbe, L.I.; Nussbaum, R.L. Mutation in the alphasynuclein gene identified in families with Parkinson's disease. *Science*, **1997**, *276*(5321), 2045-2047.
- [57] van Duijn, C.M.; Dekker, M.C.J.; Bonifati, V.; Galjaard, R.J.; Houwing-Duistermaat, J.J.; Snijders, P.J.L.M.; Testers, L.; Breedveld, G.J.; Horstink, M.; Sandkuijl, L.A.; van Swieten, J.C.; Oostra, B.A.; Heutink, P. PARK7, a Novel Locus for Autosomal Recessive Early-Onset Parkinsonism, on Chromosome 1p36. Am. J. Hum. Genet., 2001, 69(3), 629-634.
- [58] Valente, E.M.; Abou-Sleiman, P.M.; Caputo, V.; Muqit, M.M.K.; Harvey, K.; Gispert, S.; Ali, Z.; Del Turco, D.; Bentivoglio, A.R.; Healy, D.G.; Albanese, A.; Nussbaum, R.; González-Maldonado, R.; Deller, T.; Salvi, S.; Cortelli, P.; Gilks, W.P.; Latchman, D.S.; Harvey, R.J.; Dallapiccola, B.; Auburger, G.; Wood, N.W. Hereditary early-onset Parkinson's disease caused by mutations in PINK1. *Science*, 2004, 304(5674), 1158-1160.
- [59] Krüger, R.; Kuhn, W.; Müller, T.; Woitalla, D.; Graeber, M.; Kösel, S.; Przuntek, H.; Epplen, J.T.; Schöls, L.; Riess, O. Ala30Pro mutation in the gene encoding alpha-synuclein in Parkinson's disease. *Nat. Genet.*, **1998**, *18*(2), 106-108.
- [60] Spillantini, M.G.; Schmidt, M.L.; Lee, V.M.; Trojanowski, J.Q.; Jakes, R.; Goedert, M. Alpha-synuclein in Lewy bodies. *Nature*, 1997, 388(6645), 839-840.
- [61] Leroy, E.; Boyer, R.; Auburger, G.; Leube, B.; Ulm, G.; Mezey, E.; Harta, G.; Brownstein, M.J.; Jonnalagada, S.; Chernova, T.; Dehejia, A.; Lavedan, C.; Gasser, T.; Steinbach, P.J.; Wilkinson, K.D.; Polymeropoulos, M.H. The ubiquitin pathway in Parkinson's disease. *Nature*, **1998**, *395*(6701), 451-452.
- [62] Ashraf, G.M.; Bilal, N.; Suhail, N.; Hasan, S.; Banu, N. Glycosylation of purified buffalo heart galectin-1 plays crucial role in maintaining its structural and functional integrity. *Biochemistry Mosc.*, 2010, 75(12), 1450-1457.
- [63] Ashraf, G.M.; Rizvi, S.; Naqvi, S.; Suhail, N.; Bilal, N.; Hasan, S.; Tabish, M.; Banu, N. Purification, characterization, structural analysis and protein chemistry of a buffalo heart galectin-1. *Amino Acids*, **2010**, *39*(5), 1321-1332.
- [64] Ashraf, G.M.; Banu, N.; Ahmad, A.; Singh, L.P.; Kumar, R. Purification, characterization, sequencing and biological chemistry of galectin-1 purified from Capra hircus (goat) heart. *Protein J.*, 2011, 30(1), 39-51.
- [65] Buée, L.; Bussière, T.; Buée-Scherrer, V.; Delacourte, A.; Hof, P.R. Tau protein isoforms, phosphorylation and role in neurodegenerative disorders. *Brain Res. Brain Res. Rev.*, 2000, 33(1), 95-130.
- [66] Chen, H.-K.; Fernandez-Funez, P.; Acevedo, S.F.; Lam, Y.C.; Kaytor, M.D.; Fernandez, M.H.; Aitken, A.; Skoulakis, E.M.C.; Orr, H.T.; Botas, J.; Zoghbi, H.Y. Interaction of Aktphosphorylated ataxin-1 with 14-3-3 mediates neurodegeneration in spinocerebellar ataxia type 1. *Cell*, **2003**, *113*(4), 457-468.
- [67] Fujiwara, H.; Hasegawa, M.; Dohmae, N.; Kawashima, A.; Masliah, E.; Goldberg, M.S.; Shen, J.; Takio, K.; Iwatsubo, T. alpha-Synuclein is phosphorylated in synucleinopathy lesions. *Nat. Cell Biol.*, **2002**, *4*(2), 160-164.
- [68] Hasan, S.; Bilal, N.; Naqvi, S.; Ashraf, G.M.; Suhail, N.; Sharma, S.; Banu, N. Multivitamin-mineral and vitamins (E+C) supplementation modulate chronic unpredictable stress-induced oxidative damage in brain and heart of mice. *Biol. Trace Elem. Res.*, 2011, 142(3), 589-597.
- [69] Unoki, M.; Nakamura, Y. Growth-suppressive effects of BPOZ and EGR2, two genes involved in the PTEN signaling pathway. *Onco*gene, 2001, 20(33), 4457-4465.
- [70] Gary, D.S.; Mattson, M.P. PTEN regulates Akt kinase activity in hippocampal neurons and increases their sensitivity to glutamate and apoptosis. *Neuromol. Med.*, 2002, 2(3), 261-269.

- [71] Karlsson, K.E.; Novotny, M. Separation efficiency of slurry-packed liquid chromatography microcolumns with very small inner diameters. *Anal. Chem.*, **1988**, 60(17), 1662-1665.
- [72] Tao, D.; Zhang, L.; Shan, Y.; Liang, Z.; Zhang, Y. Recent advances in micro-scale and nano-scale high-performance liquidphase chromatography for proteome research. *Anal. Bioanal. Chem.*, 2011, 399(1), 229-241.
- [73] Kirsch, S.; Bindila, L. Nano-LC and HPLC-chip-ESI-MS: an emerging technique for glycobioanalysis. *Bioanalysis*, 2009, 1(7), 1307-1327.
- [74] Papac, D.I.; Shahrokh, Z. Mass spectrometry innovations in drug discovery and development. *Pharm. Res.*, 2001, 18(2), 131-145.
- [75] Siviero, A.; Bergna, M.; Famiglini, G.; Mantegazza, A.; Palma, P.; Cappiello, A. In-depth performance investigation of a nano-LC gradient generator. *Electrophoresis*, **2012**, *33*(4), 575-582.
- [76] Szekely, L.; Freitag, R. Fabrication of a versatile microanalytical system without need for clean room conditions. *Anal. Chim. Acta*, 2004, 512(1), 39-47.
- [77] Nischang, I.; Svec, F.; Frechet, J.M.J. Effect of capillary cross section geometry and size on the separation of proteins in gradient mode using monolithic poly(butyl methacrylate-co-ethylene dimethacrylate) columns. J. Chromat. A, 2009, 1216(12), 2355-2361.
- [78] Liu, Y.; Chen, L.; Sun, L. Design and Fabrication of a MEMS Flow Sensor and Its Application in Precise Liquid Dispensing. *Sensors*, 2009, 9(6), 4138-4150.
- [79] Harris, C.M. Shrinking the LC landscape. Anal. Chem., 2003, 75(3), 64A-69A.
- [80] Ericson, C.; Holm, J.; Ericson, T.; Hjertén, S. Electroosmosis- and Pressure-Driven Chromatography in Chips Using Continuous Beds. *Anal. Chem.*, 2000, 72(1), 81-87.
- [81] Murrihy, J.P.; Breadmore, M.C.; Tan, A.; McEnery, M.; Alderman, J.; O'Mathuna, C.; O'Neill, A.P.; O'Brien, P.; Advoldvic, N.; Haddad, P.R.; Glennon, J.D. Ion chromatography on-chip. J. Chromatogr. A, 2001, 924(1–2), 233-238.
- [82] Cadet, P.; Zhu, W.; Mantione, K.; Rymer, M.; Dardik, I.; Reisman, S.; Hagberg, S.; Stefano, G.B. Cyclic exercise induces antiinflammatory signal molecule increases in the plasma of Parkinson's patients. *Int. J. Mol. Med.*, **2003**, *12*(4), 485-492.
- [83] Khandurina, J.; Guttman, A. Microscale separation and analysis. *Curr. Opin. Chem. Biol.*, 2003, 7(5), 595-602.
- [84] Legido-Quigley, C.; Marlin, N.D.; Melin, V.; Manz, A.; Smith, N.W. Advances in capillary electrochromatography and micro-high performance liquid chromatography monolithic columns for separation science. *Electrophoresis*, 2003, 24(6), 917-944.
- [85] Zamfir, A.D. Recent advances in sheathless interfacing of capillary electrophoresis and electrospray ionization mass spectrometry. J. Chromatogr. A, 2007, 1159(1-2), 2-13.
- [86] Nischang, I.; Tallarek, U. Fluid dynamics in capillary and chip electrochromatography. *Electrophoresis*, 2007, 28(4), 611-626.
- [87] Fernández-la-Villa, A.; Pozo-Ayuso, D.F.; Castaño-Alvarez, M. New analytical portable instrument for microchip electrophoresis with electrochemical detection. *Electrophoresis*, **2010**, *31*(15), 2641-2649.
- [88] Phillips, T.M.; Wellner, E. Measurement of neuropeptides in clinical samples using chip-based immunoaffinity capillary electrophoresis. J. Chromatogr. A, 2006, 1111(1), 106-111.
- [89] Liu, Z.; Zhu, H.; Fang, G.G.; Walsh, K.; Mwamburi, M.; Wolozin, B.; Abdul-Hay, S.O.; Ikezu, T.; Leissring, M.A.; Qiu, W.Q. Characterization of insulin degrading enzyme and other amyloid-β degrading proteases in human serum: a role in Alzheimer's disease? J. Alzh. Dis., 2012, 29(2), 329-340.
- [90] Alper, B.J.; Schmidt, W.K. A capillary electrophoresis method for evaluation of Abeta proteolysis in vitro. J. Neurosci. Methods, 2009, 178(1), 40-45.
- [91] Vlcková, M.; Schwarz, M.A. Determination of cationic neurotransmitters and metabolites in brain homogenates by microchip electrophoresis and carbon nanotube-modified amperometry. J. Chromatogr. A, 2007, 1142(2), 214-221.
- [92] Chen, C.-M.; Chang, G.-L.; Lin, C.-H. Performance evaluation of a capillary electrophoresis electrochemical chip integrated with gold nanoelectrode ensemble working and decoupler electrodes. J. Chromatogr. A, 2008, 1194(2), 231-236.
- [93] Chicharro, M.; Arribas, A.S.; Moreno, M.; Bermejo, E.; Zapardiel, A. Comparative study of multi walled carbon nanotubes-based electrodes in micellar media and their application to micellar elec-

trokinetic capillary chromatography. *Talanta*, **2007**, *74*(3), 376-386.

- [94] Humpel, C. Identifying and validating biomarkers for Alzheimer's disease. *Trends Biotechnol.*, 2011, 29(1), 26-32.
- [95] Varesio, E.; Rudaz, S.; Krause, K.-H.; Veuthey, J.-L. Nanoscale liquid chromatography and capillary electrophoresis coupled to electrospray mass spectrometry for the detection of amyloid-β peptide related to Alzheimer's disease. J. Chromatogr. A, 2002, 974(1– 2), 135-142.
- [96] Tang, J.X.; Baranov, D.; Hammond, M.; Shaw, L.M.; Eckenhoff, M.F.; Eckenhoff, R.G. Human Alzheimer and inflammation biomarkers after anesthesia and surgery. *Anesthesiology*, 2011, 115(4), 727-732.
- [97] Misra, A.; Ganesh, S.; Shahiwala, A.; Shah, S.P. Drug delivery to the central nervous system: a review. J. Phar. Pharmac. Sci., 2003, 6(2), 252-273.
- [98] Jain, A.; Jain, A.; Gulbake, A.; Shilpi, S.; Hurkat, P.; Jain, S.K. Peptide and protein delivery using new drug delivery systems. *Crit. Rev. Ther. Drug Carrier Syst.*, 2013, 30(4), 293-329.
- [99] Rademacher, M.; Zimmerman, A.W.; Rüterjans, H.; Veerkamp, J.H.; Lücke, C. Solution structure of fatty acid-binding protein from human brain. *Mol. Cell. Biochem.*, 2002, 239(1-2), 61-68.
- [100] Thumser, A.E.; Tsai, J.; Storch, J. Collision-mediated transfer of long-chain fatty acids by neural tissue fatty acid-binding proteins (FABP): studies with fluorescent analogs. J. Mol. Neurosci., 2001, 16(2-3), 143-150.
- [101] Kumagai, A.K.; Eisenberg, J.B.; Pardridge, W.M. Absorptivemediated endocytosis of cationized albumin and a beta-endorphincationized albumin chimeric peptide by isolated brain capillaries. Model system of blood-brain barrier transport. J. Biol. Chem., 1987, 262(31), 15214-15219.
- [102] Triguero, D.; Buciak, J.L.; Pardridge, W.M. Cationization of immunoglobulin G results in enhanced organ uptake of the protein after intravenous administration in rats and primate. J. Pharmacol Exp. Ther., 1991, 258(1), 186-192.
- [103] Triguero, D.; Buciak, J.B.; Yang, J.; Pardridge, W.M. Blood-brain barrier transport of cationized immunoglobulin G: enhanced delivery compared to native protein. *Proc. Natl. Acad. Sci. U. S. A.*, **1989**, 86(12), 4761-4765.
- [104] Bickel, U.; Yoshikawa, T.; Pardridge, W.M. Delivery of peptides and proteins through the blood-brain barrier. *Adv. Drug Del. Reviews*, 2001, 46(1-3), 247-279.
- [105] Frank, H.J.; Pardridge, W.M.; Jankovic-Vokes, T.; Vinters, H.V.; Morris, W.L. Insulin binding to the blood-brain barrier in the streptozotocin diabetic rat. J. Neurochem., 1986, 47(2), 405-411.
- [106] Duffy, K.R.; Pardridge, W.M. Blood-brain barrier transcytosis of insulin in developing rabbits. *Brain Res.*, **1987**, 420(1), 32-38.
- [107] Friden, P.M.; Walus, L.R.; Musso, G.F.; Taylor, M.A.; Malfroy, B.; Starzyk, R.M. Anti-transferrin receptor antibody and antibodydrug conjugates cross the blood-brain barrier. *Proc. Natl. Acad. Sci.* U. S. A., **1991**, 88(11), 4771-4775.
- [108] Zhang, Y.; Pardridge, W.M. Conjugation of brain-derived neurotrophic factor to a blood-brain barrier drug targeting system enables neuroprotection in regional brain ischemia following intravenous injection of the neurotrophin. *Brain Res.*, 2001, 889(1-2), 49-56.
- [109] Zhang, Y.; Pardridge, W.M. Neuroprotection in transient focal brain ischemia after delayed intravenous administration of brainderived neurotrophic factor conjugated to a blood-brain barrier drug targeting system. *Stroke*, **2001**, *32*(6), 1378-1384.
- [110] Song, B.-W.; Vinters, H.V.; Wu, D.; Pardridge, W.M. Enhanced neuroprotective effects of basic fibroblast growth factor in regional brain ischemia after conjugation to a blood-brain barrier delivery vector. J. Pharmacol Exp. Ther., 2002, 301(2), 605-610.
- [111] Schwarze, S.R.; Hruska, K.A.; Dowdy, S.F. Protein transduction: unrestricted delivery into all cells? *Trends Cell Biol.*, 2000, 10(7), 290-295.
- [112] Wadia, J.S.; Dowdy, S.F. Transmembrane delivery of protein and peptide drugs by TAT-mediated transduction in the treatment of cancer. *Adv. Drug Del. Rev.*, 2005, 57(4), 579-596.
- [113] Brooks, H.; Lebleu, B.; Vivès, E. Tat peptide-mediated cellular delivery: back to basics. *Adv. Drug Del. Rev.*, 2005, 57(4), 559-577.
- [114] Derossi, D.; Joliot, A.H.; Chassaing, G.; Prochiantz, A. The third helix of the Antennapedia homeodomain translocates through biological membranes. J. Biol. Chem., 1994, 269(14), 10444-10450.

- [115] Vivès, E.; Brodin, P.; Lebleu, B. A truncated HIV-1 Tat protein basic domain rapidly translocates through the plasma membrane and accumulates in the cell nucleus. J. Biol. Chem., 1997, 272(25), 16010-16017.
- [116] Snyder, E.L.; Dowdy, S.F. Cell penetrating peptides in drug delivery. *Pharm. Res.*, 2004, 21(3), 389-393.
- [117] Kilic, U.; Kilic, E.; Dietz, G.P.H.; Bähr, M. Intravenous TAT-GDNF is protective after focal cerebral ischemia in mice. *Stroke*, 2003, 34(5), 1304-1310.
- [118] Kilic, E.; Kilic, U.; Hermann, D.M. TAT fusion proteins against ischemic stroke: current status and future perspectives. *Fro. Biosci.*, 2006, 11(1716-1721.
- [119] Dietz, G.P.H.; Valbuena, P.C.; Dietz, B.; Meuer, K.; Müller, P.; Weishaupt, J.H.; Bähr, M. Application of a blood-brain-barrierpenetrating form of GDNF in a mouse model for Parkinson's disease. *Brain Res.*, 2006, 1082(1), 61-66.
- [120] Wu, S.P.; Fu, A.L.; Wang, Y.X.; Yu, L.P.; Jia, P.Y.; Li, Q.; Jin, G.Z.; Sun, M.J. A novel therapeutic approach to 6-OHDA-induced Parkinson's disease in rats via supplementation of PTD-conjugated tyrosine hydroxylase. *Biochem. Biophys. Res. Commun.*, 2006, 346(1), 1-6.
- [121] Banks, W.A.; Lebel, C.R. Strategies for the delivery of leptin to the CNS. J. Drug Target., 2002, 10(4), 297-308.
- [122] Zlokovic, B.V. Cerebrovascular Permeability to Peptides: Manipulations of Transport Systems at the Blood-Brain Barrier. *Pharm. Res.*, **1995**, *12*(10), 1395-1406.
- [123] Pasqualini, R.; Arap, W.; McDonald, D.M. Probing the structural and molecular diversity of tumor vasculature. *Trends Mol. Med.*, 2002, 8(12), 563-571.
- [124] Torchilin, V.P. Drug targeting. Eur.J. Phar. Sci., 2000, 11 (2), S81-91.
- [125] Moghimi, S.M.; Illum, L.; Davis, S.S. Physiopathological and physicochemical considerations in targeting of colloids and drug carriers to the bone marrow. *Crit. Rev. Ther. Drug Carrier Syst.*, 1990, 7(3), 187-209.
- [126] Chavany, C.; Saison-Behmoaras, T.; Le Doan, T.; Puisieux, F.; Couvreur, P.; Hélène, C. Adsorption of oligonucleotides onto polyisohexylcyanoacrylate nanoparticles protects them against nucleases and increases their cellular uptake. *Pharm. Res.*, **1994**, *11*(9), 1370-1378.
- [127] Calvo, P.; Gouritin, B.; Chacun, H.; Desmaële, D.; D'Angelo, J.; Noel, J.-P.; Georgin, D.; Fattal, E.; Andreux, J.P.; Couvreur, P. Long-Circulating PEGylated Polycyanoacrylate Nanoparticles as New Drug Carrier for Brain Delivery. *Pharm. Res.*, **2001**, *18*(8), 1157-1166.
- [128] Ashraf, G.M.; Perveen, A.; Zaidi, S.K.; Tabrez, S.; Kamal, M.A.; Banu, N. Studies on the role of goat heart galectin-1 as an erythrocyte membrane perturbing agent. *Saudi J. Biol. Sci.*, (article in press).
- [129] Ashraf, G.M.; Perveen, A.; Tabrez, S.; Zaidi, S.K.; Kamal, M.A.; Banu, N. Studies on the role of goat heart galectin-1 as a tool for detecting post-malignant changes in glycosylation pattern. *Saudi J. Biol. Sci.*, (article in press).
- [130] Ashraf, G.M. Galectins: A Research Overview. LAP LAMBERT Academic Publishing, 2011.
- [131] Godard, G.; Boutorine, A.S.; Saison-Behmoaras, E.; Hélène, C. Antisense Effects of Cholesterol-Oligodeoxynucleotide Conjugates Associated with Poly(alkylcyanoacrylate) Nanoparticles. *Eur. J. Biochem.*, **1995**, *232*(2), 404-410.
- [132] Fattal, E.; Vauthier, C.; Aynie, I.; Nakada, Y.; Lambert, G.; Malvy, C.; Couvreur, P. Biodegradable polyalkylcyanoacrylate nanoparticles for the delivery of oligonucleotides. *J. Control. Release*, 1998, 53(1-3), 137-143.
- [133] Torchilin, V.P. Affinity liposomes *in vivo*: factors influencing target accumulation. J. Mol. Recog., **1996**, 9(5-6), 335-346.
- [134] Allen, T.M. Liposomes. Opportunities in drug delivery. Drugs, 1997, 54(4), 8-14.
- [135] Kwon, G.S.; Okano, T. Soluble Self-Assembled Block Copolymers for Drug Delivery. *Pharm. Res.*, **1999**, *16*(5), 597-600.
- [136] Alakhov, V.; Klinski, E.; Lemieux, P.; Pietrzynski, G.; Kabanov, A. Block copolymeric biotransport carriers as versatile vehicles for drug delivery. *Expert Opin. Biol. Ther.*, 2001, 1(4), 583-602.
- [137] Torchilin, V.P. PEG-based micelles as carriers of contrast agents for different imaging modalities. *Adv. Drug Del. Rev.*, 2002, 54(2), 235-252.

- [138] Kabanov, A.V.; Alakhov, V.Y. Pluronic block copolymers in drug delivery: from micellar nanocontainers to biological response modifiers. *Crit. Rev. Ther. Drug Carrier Syst.*, 2002, 19(1), 1-72.
- [139] Torchilin, V.P. Polymer-coated long-circulating microparticulate pharmaceuticals. *J. Microencapsul.*, **1998**, *15*(1), 1-19.
- [140] Gref, R.; Minamitake, Y.; Peracchia, M.T.; Trubetskoy, V.; Torchilin, V.; Langer, R. Biodegradable long-circulating polymeric nanospheres. *Science*, **1994**, *263*(5153), 1600-1603.
- [141] Stolnik, S.; Dunn, S.E.; Garnett, M.C.; Davies, M.C.; Coombes, A.G.; Taylor, D.C.; Irving, M.P.; Purkiss, S.C.; Tadros, T.F.; Davis, S.S. Surface modification of poly(lactide-co-glycolide) nanospheres by biodegradable poly(lactide)-poly(ethylene glycol) copolymers. *Pharm. Res.*, **1994**, *11*(12), 1800-1808.
- [142] Peracchia, M.T.; Vauthier, C.; Desmaële, D.; Gulik, A.; Dedieu, J.C.; Demoy, M.; d'Angelo, J.; Couvreur, P. Pegylated nanoparticles from a novel methoxypolyethylene glycol cyanoacrylatehexadecyl cyanoacrylate amphiphilic copolymer. *Pharm. Res.*, **1998**, 15(4), 550-556.
- [143] Zhang, Y.; Boado, R.J.; Pardridge, W.M. Marked enhancement in gene expression by targeting the human insulin receptor. J. Gene Med., 2003, 5(2), 157-163.
- [144] Zhang, Y.; Jeong Lee, H.; Boado, R.J.; Pardridge, W.M. Receptormediated delivery of an antisense gene to human brain cancer cells. *J. Gene Med.*, 2002, 4(2), 183-194.
- [145] Pardridge, W.M. The Blood-Brain Barrier: Bottleneck in Brain Drug Development. *Neurorx*, **2005**, *2*(1), 3-14.
- [146] Shi, N.; Zhang, Y.; Zhu, C.; Boado, R.J.; Pardridge, W.M. Brainspecific expression of an exogenous gene after i.v. administration. *PNAS*, 2001, 98(22), 12754-12759.
- [147] Pardridge, W.M. Drug and gene delivery to the brain: the vascular route. *Neuron*, 2002, 36(4), 555-558.
- [148] Vinogradov, S.; Batrakova, E.; Kabanov, A. Poly(ethylene glycol)– polyethyleneimine NanoGelTM particles: novel drug delivery systems for antisense oligonucleotides. *Colloids and Surfaces B: Biointerfaces*, 1999, 16(1-4), 291-304.
- [149] Bronich, T.K.; Bontha, S.; Shlyakhtenko, L.S.; Bromberg, L.; Hatton, T.A.; Kabanov, A.V. Template-assisted synthesis of nanogels from Pluronic-modified poly(acrylic acid). J. Drug Target., 2006, 14(6), 357-366.
- [150] Bontha, S.; Kabanov, A.V.; Bronich, T.K. Polymer micelles with cross-linked ionic cores for delivery of anticancer drugs. J. Control. Release, 2006, 114(2), 163-174.
- [151] Vinogradov, S.V.; Zeman, A.D.; Batrakova, E.V.; Kabanov, A.V. Polyplex Nanogel formulations for drug delivery of cytotoxic nucleoside analogs. J. Control. Release, 2005, 107(1), 143-157.
- [152] Gozes, I. Neuroprotective peptide drug delivery and development: potential new therapeutics. *Trends Neurosci.*, 2001, 24(12), 700-705.
- [153] Kroll, R.A.; Neuwelt, E.A. Outwitting the blood-brain barrier for therapeutic purposes: osmotic opening and other means. *Neurosurgery*, **1998**, 42(5), 1083-1099; discussion 1099-1100.
- [154] Desnick, R.J.; Schuchman, E.H. Enzyme replacement and enhancement therapies: lessons from lysosomal disorders. *Nat. Rev. Genet.*, 2002, 3(12), 954-966.
- [155] Downs-Kelly, E.; Jones, M.Z.; Alroy, J.; Cavanagh, K.T.; King, B.; Lucas, R.E.; Baker, J.C.; Kraemer, S.A.; Hopwood, J.J. Caprine mucopolysaccharidosis IIID: a preliminary trial of enzyme replacement therapy. J. Mol. Neurosci., 2000, 15(3), 251-262.
- [156] Banks, W.A. Is obesity a disease of the blood-brain barrier? Physiological, pathological, and evolutionary considerations. *Curr. Pharm. Des.*, **2003**, *9*(10), 801-809.
- [157] Pardridge, W.M. Recent developments in peptide drug delivery to the brain. *Pharmacol. Toxicol.*, **1992**, *71*(1), 3-10.
- [158] Kabanov, A.V.; Levashov, A.V.; Martinek, K. Transformation of Water-Soluble Enzymes into Membrane Active Form by Chemical Modification. Ann. N. Y. Acad. Sci., 1987, 501(1), 63-66.
- [159] Kabanov, A.V.; Levashov, A.V.; Alakhov, V. Lipid modification of proteins and their membrane transport. *Protein Eng.*, **1989**, *3*(1), 39-42.
- [160] Hashimoto, M.; Takada, K.; Kiso, Y.; Muranishi, S. Synthesis of palmitoyl derivatives of insulin and their biological activities. *Pharm. Res.*, **1989**, 6(2), 171-176.
- [161] Colsky, A.S.; Peacock, J.S. Palmitate-derivatized antibodies can specifically "arm" macrophage effector cells for ADCC. *J. Leukoc. Biol.*, **1991**, *49*(1), 1-7.

- [162] Kabanov, A.V.; Ovcharenko, A.V.; Melik-Hubarov, N.S.; Bannikov, A.I.; Alakhov, V.; Kiselev, V.I.; Sveshnikov, P.G.; Kiselev, O.I.; Levashov, A.V.; Severin, E.S. Fatty acid acylated antibodies against virus suppress its reproduction in cells. *FEBS Lett.*, **1989**, 250(2), 238-240.
- [163] Kabanov, A.V.; Kabanov, V.A. DNA Complexes with Polycations for the Delivery of Genetic Material into Cells. *Bioconjug. Chem.*, 1995, 6(1), 7-20.
- [164] Alakhov, V.; Kabanov, A.V.; Batrakova, E.V.; Koromyslova, I.A.; Levashov, A.V.; Severin, E.S. Increasing cytostatic effects of ricin A chain and Staphylococcus aureus enterotoxin A through *in vitro* hydrophobization with fatty acid residues. *Biotechnol. Appl. Biochem.*, **1990**, *12*(1), 94-98.
- [165] Chekhonin, V.P.; Kabanov, A.V.; Zhirkov, Y.A.; Morozov, G.V. Fatty acid acylated Fab-fragments of antibodies to neurospecific proteins as carriers for neuroleptic targeted delivery in brain. *FEBS Lett.*, **1991**, 287(1-2), 149-152.
- [166] Robert, S.; Domurado, D.; Thomas, D.; Chopineau, J. Fatty acid acylation of RNase A using reversed micelles as microreactors. *Biochem. Biophys. Res. Commun.*, 1993, 196(1), 447-454.
- [167] Robert, S.; Domurado, D.; Thomas, D.; Chopineau, J. Optimization of RNase A Artificial Hydrophobization in AOT Reversed Micelles. Ann. N. Y. Acad. Sci., 1995, 750(1), 121-124.
- [168] Slepnev, V.I.; Phalente, L.; Labrousse, H.; Melik-Nubarov, N.S.; Mayau, V.; Goud, B.; Buttin, G.; Kabanov, A.V. Fatty acid acylated peroxidase as a model for the study of interactions of hydrophobically-modified proteins with mammalian cells. *Bioconjug. Chem.*, **1995**, *6*(5), 608-615.
- [169] Chekhonin, V.P.; Ryabukhin, I.A.; Zhirkov, Y.A.; Kashparov, I.A.; Dmitriyeva, T.B. Transport of hydrophobized fragments of antibodies through the blood-brain barrier. *Neuroreport*, **1995**, 7(1), 129-132.
- [170] Chopineau, J.; Robert, S.; Fénart, L.; Cecchelli, R.; Lagoutte, B.; Paitier, S.; Dehouck, M.P.; Domurado, D. Monoacylation of ribonuclease A enables its transport across an *in vitro* model of the blood-brain barrier. *J. Control. Release*, **1998**, *56*(1-3), 231-237.
- [171] Batrakova, E.V.; Vinogradov, S.V.; Robinson, S.M.; Niehoff, M.L.; Banks, W.A.; Kabanov, A.V. Polypeptide point modifications with fatty acid and amphiphilic block copolymers for enhanced brain delivery. *Bioconjug. Chem.*, 2005, 16(4), 793-802.
- [172] Miller, D.W.; Batrakova, E.V.; Waltner, T.O.; Alakhov, V.; Kabanov, A.V. Interactions of pluronic block copolymers with brain microvessel endothelial cells: evidence of two potential pathways for drug absorption. *Bioconjug. Chem.*, **1997**, *8*(5), 649-657.
- [173] Batrakova, E.V.; Li, S.; Vinogradov, S.V.; Alakhov, V.Y.; Miller, D.W.; Kabanov, A.V. Mechanism of pluronic effect on Pglycoprotein efflux system in blood-brain barrier: contributions of energy depletion and membrane fluidization. *J. Pharmacol. Exp. Ther.*, **2001**, 299(2), 483-493.
- [174] Buzea, C.; Pacheco, I.I.; Robbie, K. Nanomaterials and nanoparticles: Sources and toxicity. *Biointerphases*, 2007, 2(4), MR17-MR71.
- [175] Pollak, P.; Fraix, V.; Krack, P.; Moro, E.; Mendes, A.; Chabardes, S.; Koudsie, A.; Benabid, A.-L. Treatment results: Parkinson's disease. *Movement disorders*, 2002, *17 Suppl 3*(S75-83.
- [176] Freed, C.R.; Greene, P.E.; Breeze, R.E.; Tsai, W.-Y.; DuMouchel, W.; Kao, R.; Dillon, S.; Winfield, H.; Culver, S.; Trojanowski, J.Q.; Eidelberg, D.; Fahn, S. Transplantation of Embryonic Dopamine Neurons for Severe Parkinson's Disease. *N. Engl. J. Med.*, **2001**, 344(10), 710-719.
- [177] Dickson, D.W. Misfolded, protease-resistant proteins in animal models and human neurodegenerative disease. J. Clin. Investigation, 2002, 110(10), 1403-1405.
- [178] Woodruff, B.K.; Graff-Radford, N.R.; Ferman, T.J.; Dickson, D.W.; DeLucia, M.W.; Crook, J.E.; Arvanitakis, Z.; Brassler, S.; Waters, C.; Barker, W.; Duara, R. Family history of dementia is a risk factor for Lewy body disease. *Neurology*, **2006**, *66*(12), 1949-1950.
- [179] Müller, R.H.; Keck, C.M. Drug delivery to the brain--realization by novel drug carriers. J. Nanosci. Nanotechnol., 2004, 4(5), 471-483.
- [180] Hyuk Im, S.; Jeong, U.; Xia, Y. Polymer hollow particles with controllable holes in their surfaces. *Nat. Mater.*, 2005, 4(9), 671-675.
- [181] Oishi, M.; Hayashi, H.; Iijima, M.; Nagasaki, Y. Endosomal release and intracellular delivery of anticancer drugs using pH-sensitive PEGylated nanogels. J. Mater. Chem., 2007, 17(35), 3720-3725.

- [182] Balogh, L.P. Why do we have so many definitions for nanoscience and nanotechnology? *Nanomedicine Nanotech., Biol. Med.*, 2010, 6(3), 397-398.
- [183] Shephard, M.J.; Todd, D.; Adair, B.M.; Po, A.L.W.; Mackie, D.P.; Scott, E.M. Immunogenicity of bovine parainfluenza type 3 virus proteins encapsulated in nanoparticle vaccines, following intranasal administration to mice. *Res. Vet. Sci.*, **2003**, *74*(2), 187-190.
- [184] Cui, Z.; Mumper, R.J. Intranasal administration of plasmid DNAcoated nanoparticles results in enhanced immune responses. J. Phar. Pharmacol., 2002, 54(9), 1195-1203.
- [185] Vijayanathan, V.; Thomas, T.; Thomas, T.J. DNA nanoparticles and development of DNA delivery vehicles for gene therapy. *Biochemistry (Mosc.)*, 2002, 41(48), 14085-14094.
- [186] Cleland, J.L. Solvent evaporation processes for the production of controlled release biodegradable microsphere formulations for therapeutics and vaccines. *Biotechnol. Prog.*, **1998**, *14*(1), 102-107.
- [187] Aukunuru, J.V.; Ayalasomayajula, S.P.; Kompella, U.B. Nanoparticle formulation enhances the delivery and activity of a vascular endothelial growth factor antisense oligonucleotide in human retinal pigment epithelial cells. J. Phar. Pharmacol., 2003, 55(9), 1199-1206.
- [188] Desai, N. Challenges in development of nanoparticle-based therapeutics. AAPS J., 2012, 14(2), 282-295.
- [189] Liu, Y. Review on nano-drugs. Natural Sci., 2010, 02(01), 41-48.
- [190] Singh, M.N.; Hemant, K.S.Y.; Ram, M.; Shivakumar, H.G. Microencapsulation: A promising technique for controlled drug delivery. *Res. Pharm. Sci.*, **2010**, *5*(2), 65-77.
- [191] Masaro, L.; Zhu, X.X. Physical models of diffusion for polymer solutions, gels and solids. *Prog. Polyr. Sci.*, **1999**, *24*(5), 731-775.
- [192] Hong, X.; Wei, L.; Ma, L.; Chen, Y.; Liu, Z.; Yuan, W. Novel preparation method for sustained-release PLGA microspheres using water-in-oil-in-hydrophilic-oil-in-water emulsion. *Int. J. Nanomedicine.*, 2013, 8, 2433-2441.
- [193] Patomchaiviwat, V.; Paeratakul, O.; Kulvanich, P. Formation of Inhalable Rifampicin–Poly(l-lactide) Microparticles by Supercritical Anti-solvent Process. AAPS PharmSciTech., 2008, 9(4), 1119-1129.
- [194] Coelho, J.F.; Ferreira, P.C.; Alves, P.; Cordeiro, R.; Fonseca, A.C.; Gois, J.R.; Gil, M.H. Drug delivery systems: Advanced technologies potentially applicable in personalized treatments. *EPMA J.*, **2010**, *1*(1), 164-209.
- [195] Fu, Y.; Kao, W.J. Drug Release Kinetics and Transport Mechanisms of Non-degradable and Degradable Polymeric Delivery Systems. *Expert Opin. Drug Deliv.*, 2010, 7(4), 429-444.
- [196] De Jong, W.H.; Borm, P.J.A. Drug delivery and nanoparticles: Applications and hazards. Int. J. Nanomedicine, 2008, 3(2), 133-149.
- [197] Frank, A.; Rath, S.K.; Venkatraman, S.S. Controlled release from bioerodible polymers: effect of drug type and polymer composition. J. Control. Release, 2005, 102(2), 333-344.
- [198] Richardson, S.C.; Kolbe, H.V.; Duncan, R. Potential of low molecular mass chitosan as a DNA delivery system: biocompatibility, body distribution and ability to complex and protect DNA. *Int. J. Pharm.*, **1999**, *178*(2), 231-243.
- [199] Kim, J.S.; Yoon, T.-J.; Yu, K.N.; Kim, B.G.; Park, S.J.; Kim, H.W.; Lee, K.H.; Park, S.B.; Lee, J.-K.; Cho, M.H. Toxicity and tissue distribution of magnetic nanoparticles in mice. *Toxicol. Sci.*, 2006, 89(1), 338-347.
- Yang, S.-t.; Guo, W.; Lin, Y.; Deng, X.-y.; Wang, H.-f.; Sun, H.-f.; Liu, Y.-f.; Wang, X.; Wang, W.; Chen, M.; Huang, Y.-p.; Sun, Y.-P. Biodistribution of Pristine Single-Walled Carbon Nanotubes *In vivo*†. *J. Phys. Chem. C*, 2007, *111*(48), 17761-17764.
- [201] Lam, C.-W.; James, J.T.; McCluskey, R.; Hunter, R.L. Pulmonary Toxicity of Single-Wall Carbon Nanotubes in Mice 7 and 90 Days After Intratracheal Instillation. *Toxicol. Sci.*, 2004, 77(1), 126-134.
- [202] Service, R.F. Nanomaterials Show Signs of Toxicity. Science, 2003, 300(5617), 243-243.
- [203] Kasemets, K.; Ivask, A.; Dubourguier, H.-C.; Kahru, A. Toxicity of nanoparticles of ZnO, CuO and TiO2 to yeast Saccharomyces cerevisiae. *Toxicol. In vitro*, 2009, 23(6), 1116-1122.
- [204] Hussain, S.M.; Hess, K.L.; Gearhart, J.M.; Geiss, K.T.; Schlager, J.J. *In vitro* toxicity of nanoparticles in BRL 3A rat liver cells. *Toxicol. In vitro*, 2005, 19(7), 975-983.
- [205] Aruoja, V.; Dubourguier, H.-C.; Kasemets, K.; Kahru, A. Toxicity of nanoparticles of CuO, ZnO and TiO2 to microalgae Pseu-

dokirchneriella subcapitata. Sci. Total Environ., 2009, 407(4), 1461-1468.

- [206] Pirson, P.; Steiger, R.; Trouet, A. The disposition of free and liposomally encapsulated antimalarial primaquine in mice. *Biochem. Pharmacol.*, **1982**, *31*(21), 3501-3507.
- [207] Yang, M.-H.; Lin, C.-H.; Chang, L.W.; Lin, P. Application of ICP-MS for the study of disposition and toxicity of metal-based nanomaterials. *Meth. Mol. Biol.*, 2012, 926(345-359.
- [208] Stern, S.T.; McNeil, S.E. Nanotechnology safety concerns revisited. *Toxicol. Sci.*, 2008, 101(1), 4-21.
- [209] Modi, G.; Pillay, V.; Choonara, Y.E. Advances in the treatment of neurodegenerative disorders employing nanotechnology. *Ann. N. Y. Acad. Sci.*, 2010, 1184, 154-172.
- [210] Bernardi, A.; Frozza, R.L.; Horn, A.P.; Campos, M.M.; Calixto, J.B.; Salbego, C.; Pohlmann, A.R.; Guterres, S.S.; Battastini, A.M.O. Protective effects of indomethacin-loaded nanocapsules against oxygen-glucose deprivation in organotypic hippocampal slice cultures: involvement of neuroinflammation. *Neurochem. Int.*, 2010, 57(6), 629-636.
- [211] Modi, G.; Pillay, V.; Choonara, Y.E.; Ndesendo, V.M.K.; du Toit, L.C.; Naidoo, D. Nanotechnological applications for the treatment of neurodegenerative disorders. *Prog. Neurobiol.*, 2009, 88(4), 272-285.

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- [212] Wang, T.; Wang, C.-Y.; Shan, Z.-Y.; Teng, W.-P.; Wang, Z.-Y. Clioquinol reduces zinc accumulation in neuritic plaques and inhibits the amyloidogenic pathway in AβPP/PS1 transgenic mouse brain. J. Alzh. Dis., 2012, 29(3), 549-559.
- [213] Cui, Z.; Lockman, P.R.; Atwood, C.S.; Hsu, C.-H.; Gupte, A.; Allen, D.D.; Mumper, R.J. Novel D-penicillamine carrying nanoparticles for metal chelation therapy in Alzheimer's and other CNS diseases. *Eur. J. Phar. Biopharmac.*, **2005**, *59*(2), 263-272.
- [214] Matsumoto, K.; Sato, C.; Naka, Y.; Whitby, R.; Shimizu, N. Stimulation of neuronal neurite outgrowth using functionalized carbon nanotubes. *Nanotech.*, 2010, 21(11).
- [215] Yang, Z.; Zhang, Y.; Yang, Y.; Sun, L.; Han, D.; Li, H.; Wang, C. Pharmacological and toxicological target organelles and safe use of single-walled carbon nanotubes as drug carriers in treating Alzheimer disease. *Nanomedicine.*, 2010, 6(3), 427-441.
- [216] Pai, A.S.; Rubinstein, I.; Onyüksel, H. PEGylated phospholipid nanomicelles interact with beta-amyloid((1-42)) and mitigate its beta-sheet formation, aggregation and neurotoxicity *in vitro*. *Peptides*, 2006, 27(11), 2858-2866.
- [217] Mourtas, S.; Canovi, M.; Zona, C.; Aurilia, D.; Niarakis, A.; La Ferla, B.; Salmona, M.; Nicotra, F.; Gobbi, M.; Antimisiaris, S.G. Curcumin-decorated nanoliposomes with very high affinity for amyloid-β1-42 peptide. *Biomaterials*, 2011, 32(6), 1635-1645.